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# Field asymmetric waveform ion mobility spectrometry combined with mass spectrometry for the analysis of anabolic steroids

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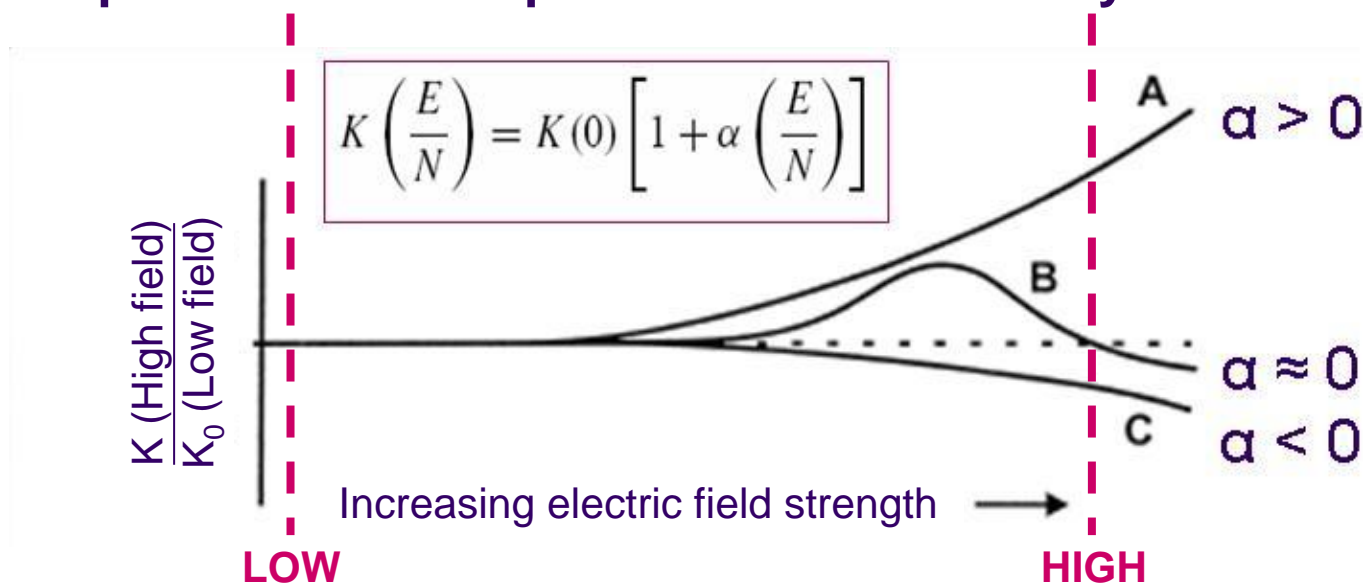
# Outline

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- Aims
- What is FAIMS?
- FAIMS-MS and LC-FAIMS-MS instrumentation
- FAIMS-MS separation of steroid metabolites – sulfates and glucuronides
- LC-FAIMS-MS analysis of steroids in urine matrix

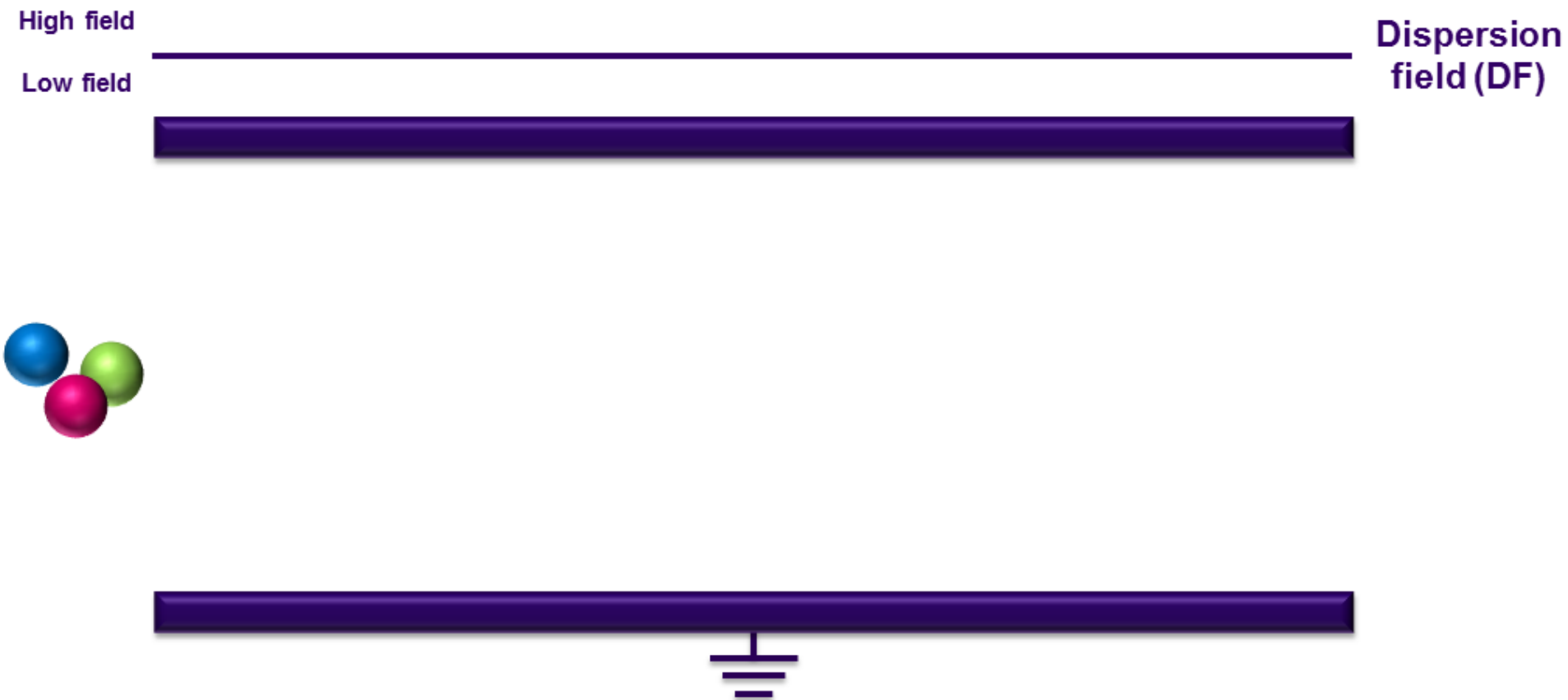
# What is FAIMS?

- Field asymmetric waveform ion mobility spectrometry
- Separation of ions based upon their non-linear relationship between mobility and increasing electric field strength
- **Ion separation based upon differential mobility**

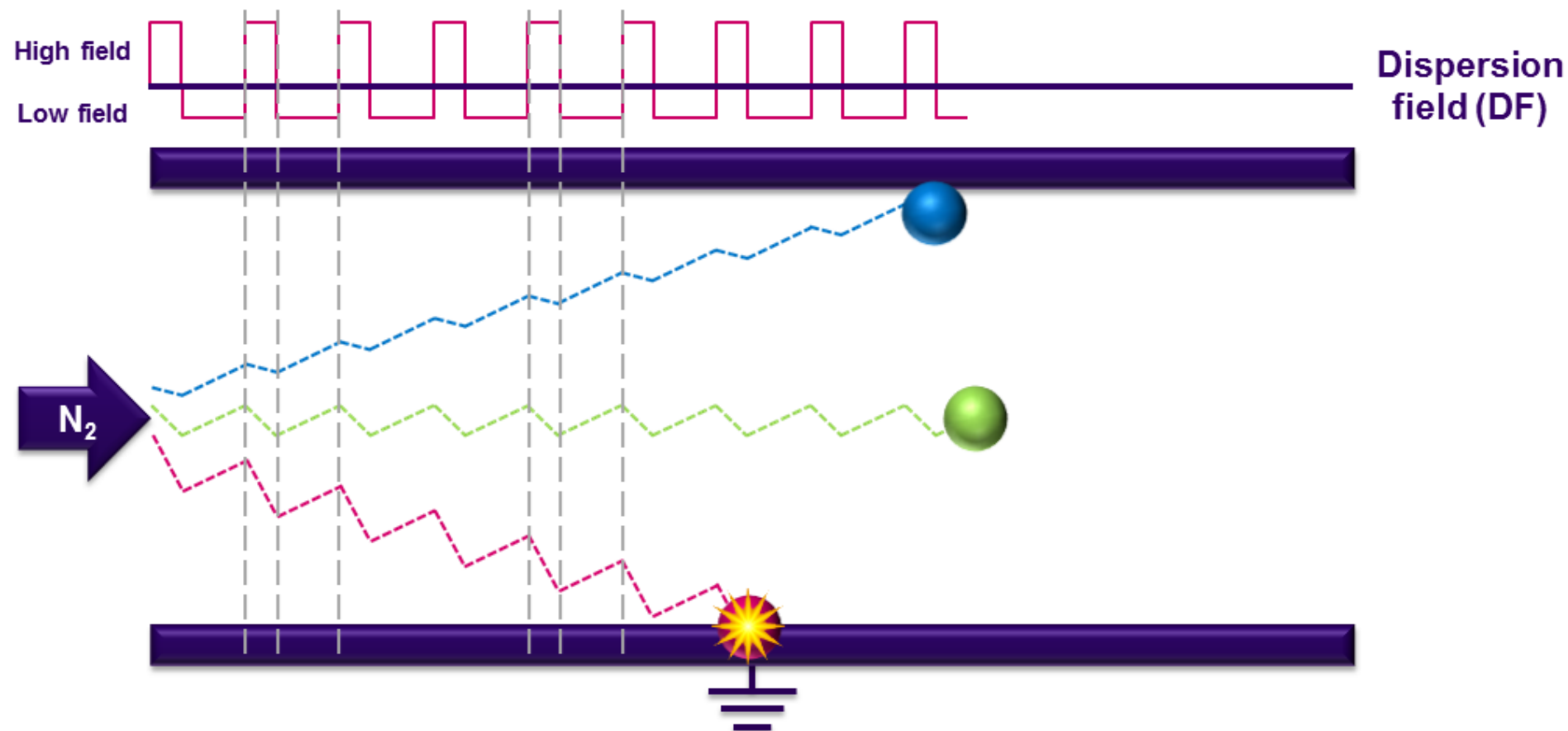


[Purves R W, Guevremont R, Anal. Chem. 1999, 71, 2346-2357]

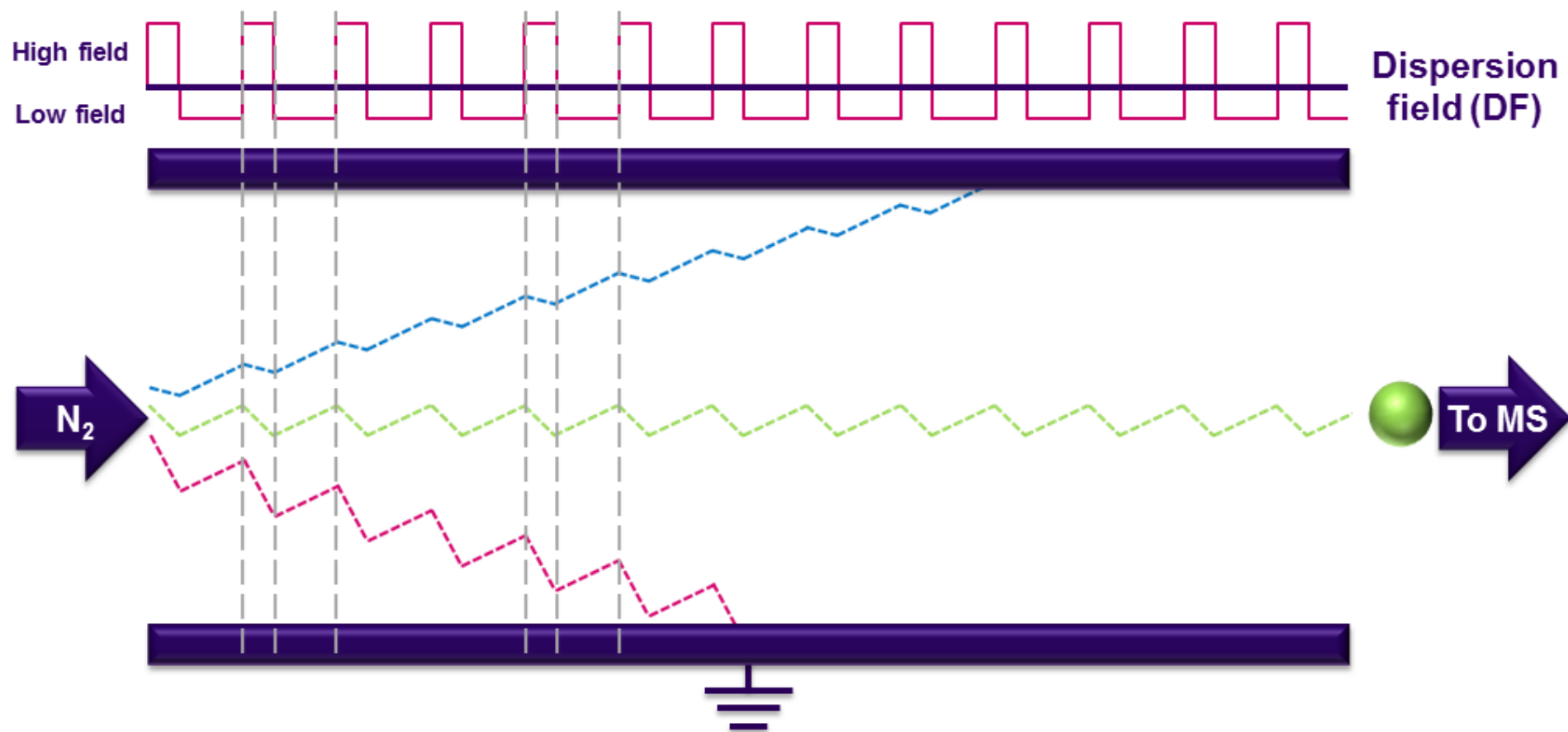
# What is FAIMS?



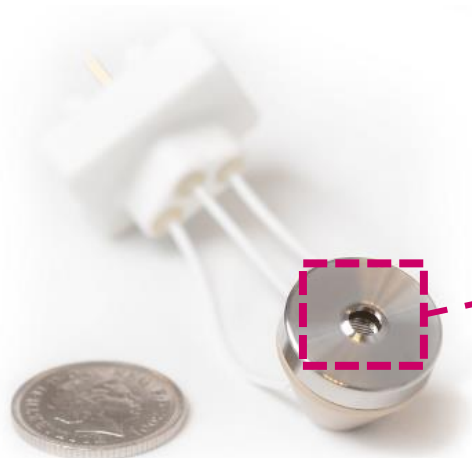
# What is FAIMS?



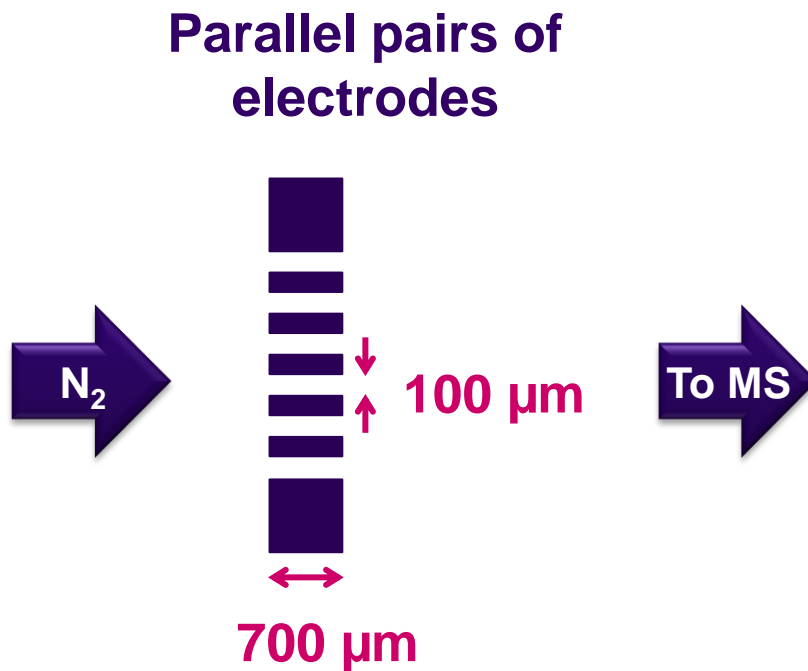
# What is FAIMS?



# Owlstone chip-based FAIMS

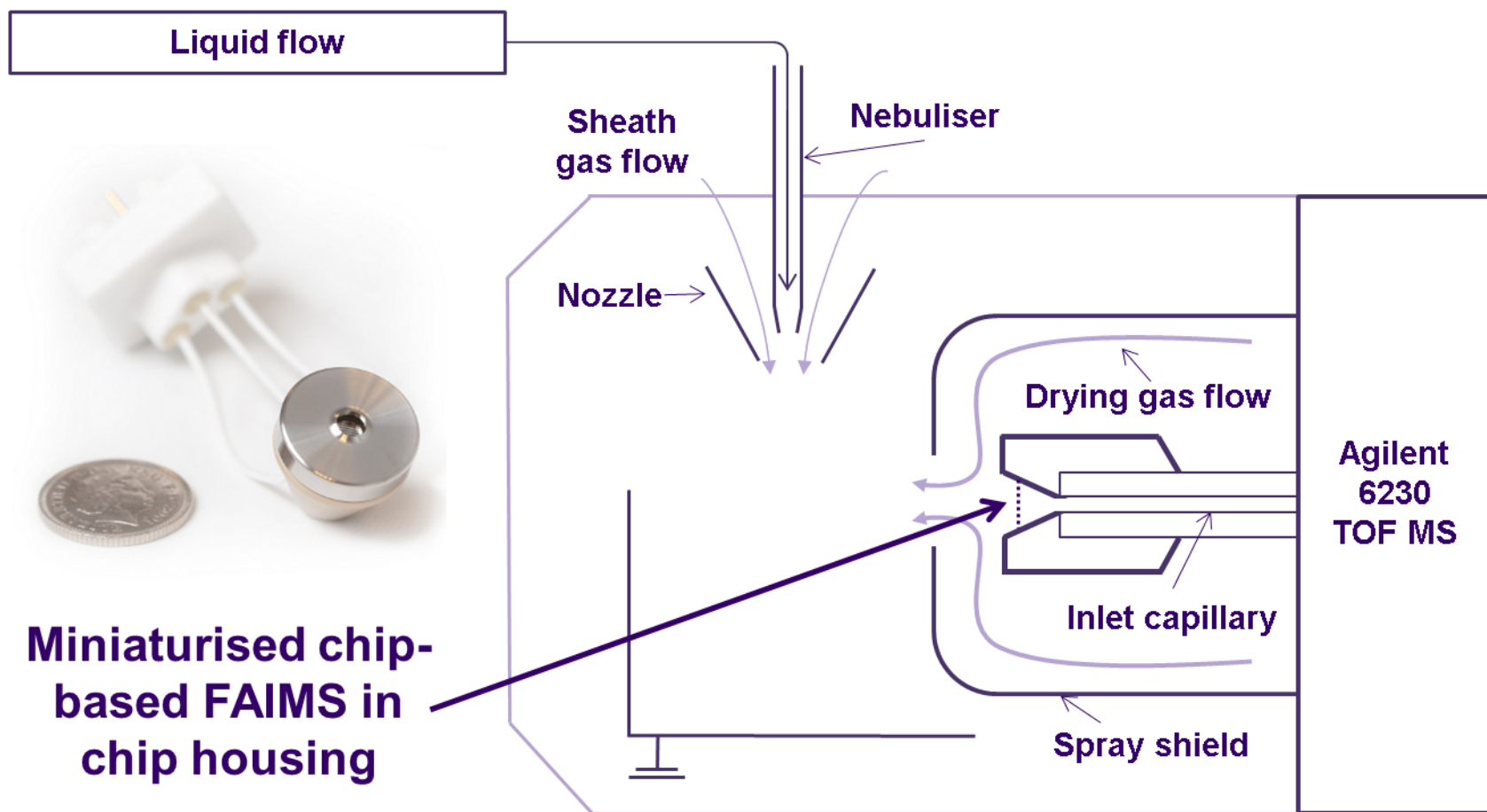


Miniaturised chip-based FAIMS in chip housing



Side view of FAIMS chip

# Owlstone chip-based FAIMS



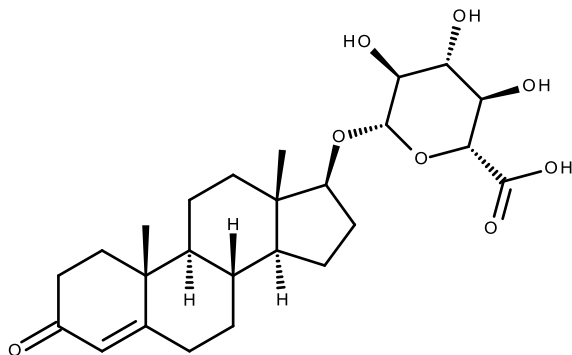


# Targeted steroid compounds

Testosterone  
glucuronide

**TG**

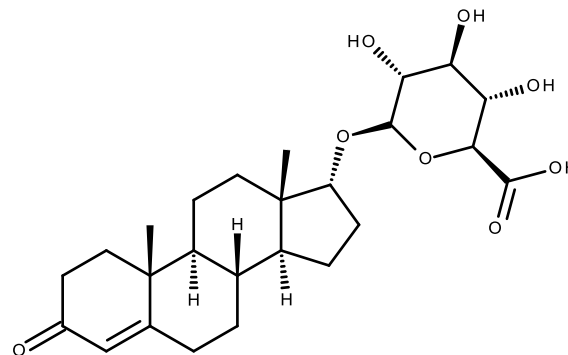
MW 464



Epitestosterone  
glucuronide

**ETG**

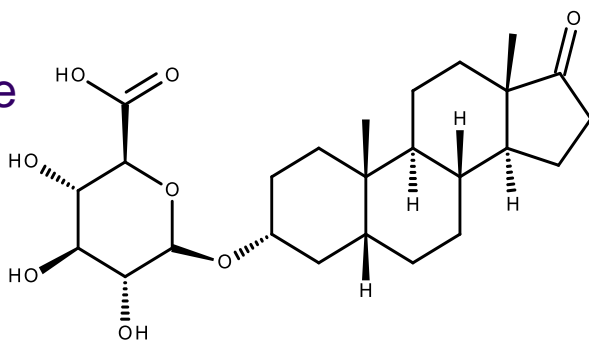
MW 464



Etiocholanolone  
glucuronide

**ECG**

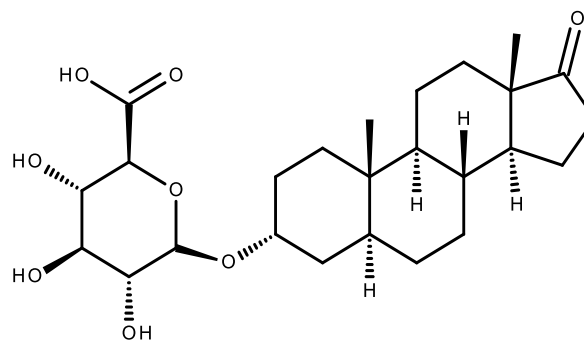
MW 466



Androsterone  
glucuronide

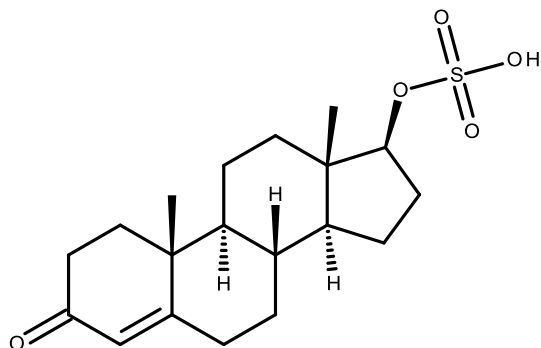
**ADG**

MW 466

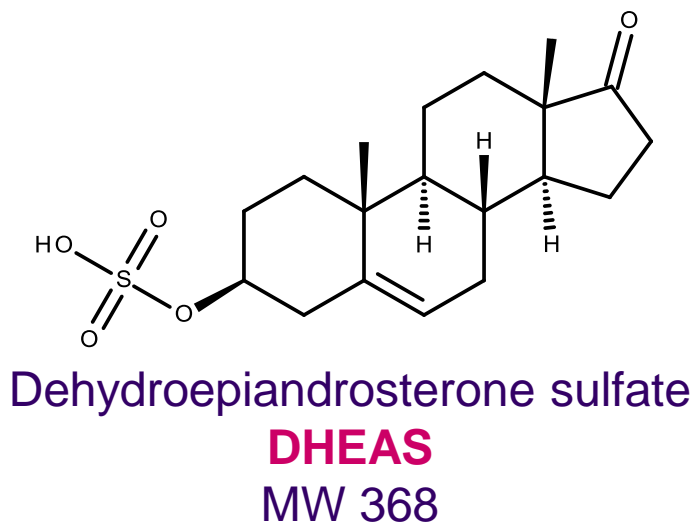
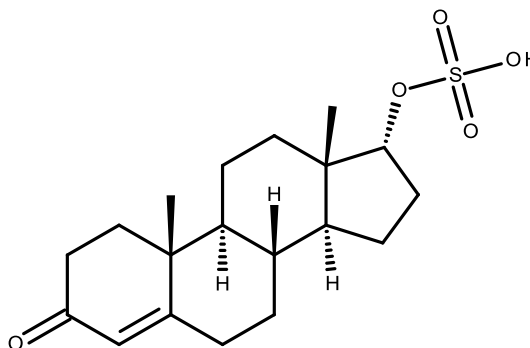


# Targeted steroid compounds

Testosterone  
sulfate  
**TS**  
MW 368



Epitestosterone  
sulfate  
**ETS**  
MW 368



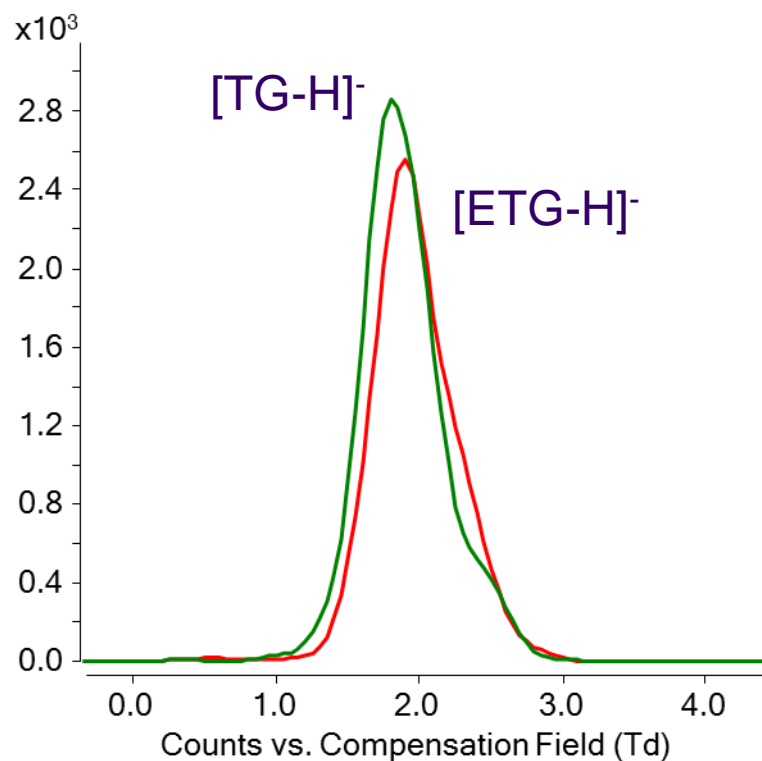
# Separation of steroids using MS alone

| <i>m/z</i> | ETG | TG | ECG | ADG | DHEAS | TS | ETS |
|------------|-----|----|-----|-----|-------|----|-----|
| ETG        |     |    |     |     |       |    |     |
| TG         |     |    |     |     |       |    |     |
| ECG        |     |    |     |     |       |    |     |
| ADG        |     |    |     |     |       |    |     |
| DHEAS      |     |    |     |     |       |    |     |
| TS         |     |    |     |     |       |    |     |
| ETS        |     |    |     |     |       |    |     |

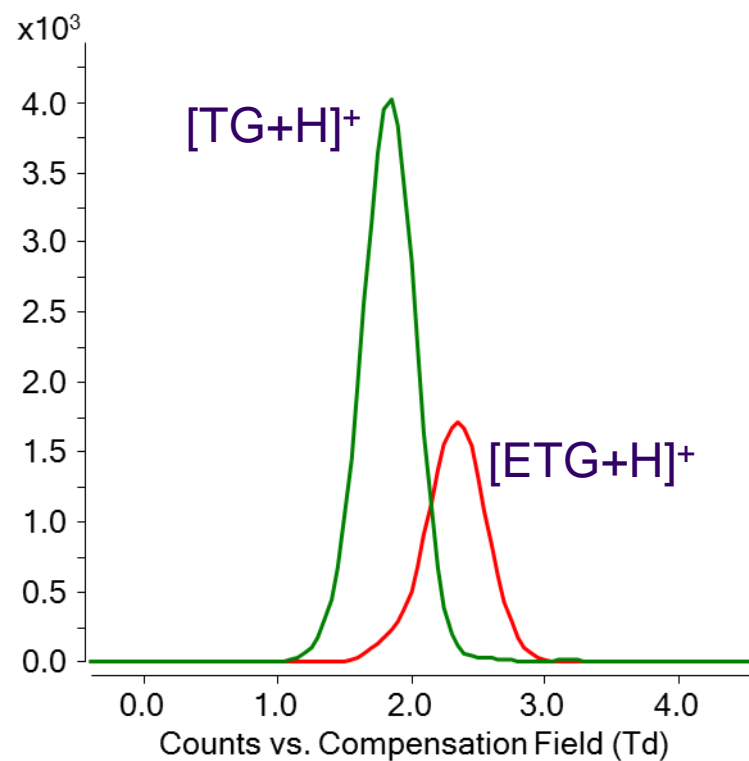
Isobaric pairs/trio unresolved using MS alone

# FAIMS-MS of steroid metabolites

## Negative ion mode



## Positive ion mode



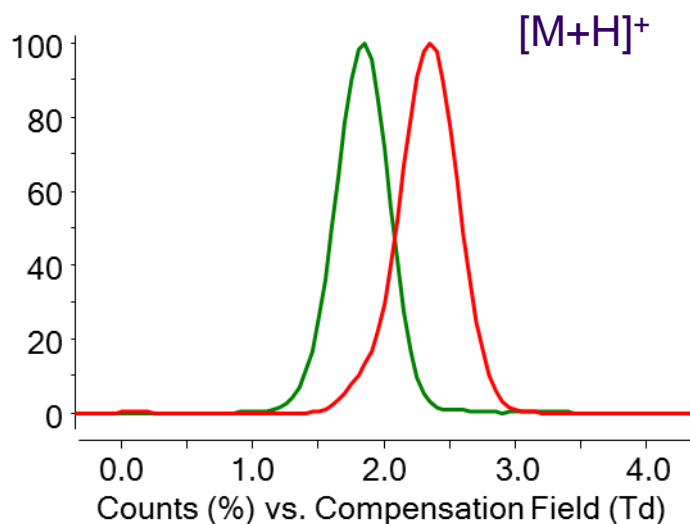
Dispersion Field 300 Td

Initial FAIMS-MS in MeOH:H<sub>2</sub>O 50:50 + 0.1% FA

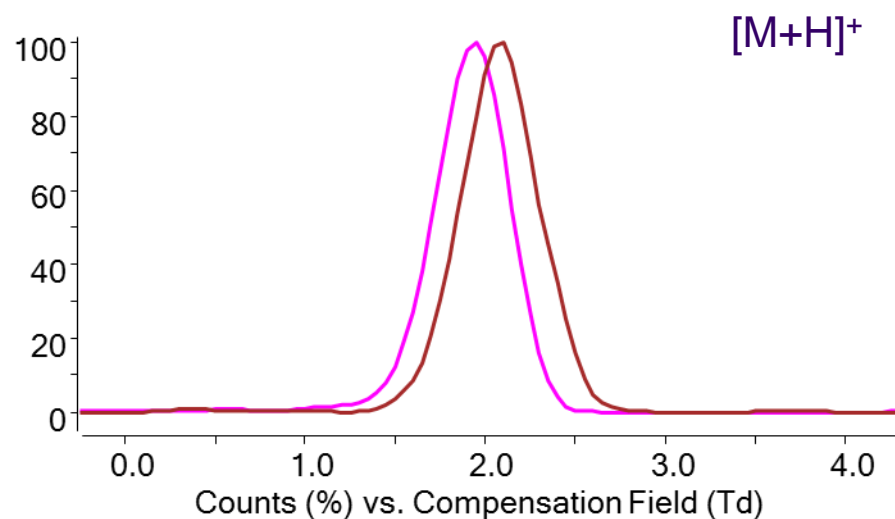
# FAIMS-MS of steroid metabolites

## Positive ion mode

Dispersion Field 300 Td



Testosterone glucuronide  
Epitestosterone glucuronide

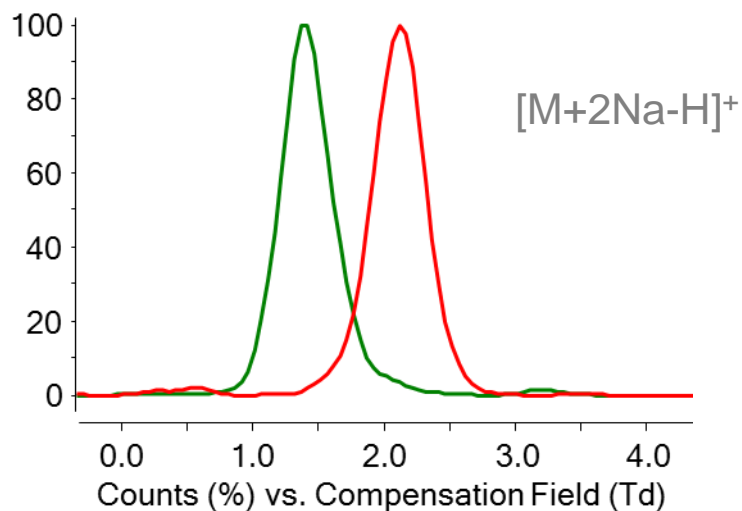


Testosterone sulfate  
Epitestosterone sulfate

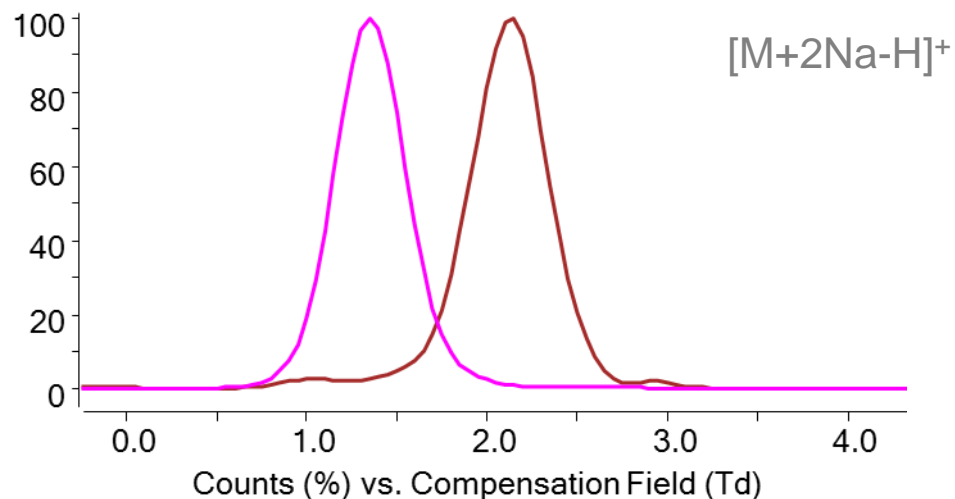
# FAIMS-MS of steroid metabolites

## Positive ion mode

Dispersion Field 300 Td

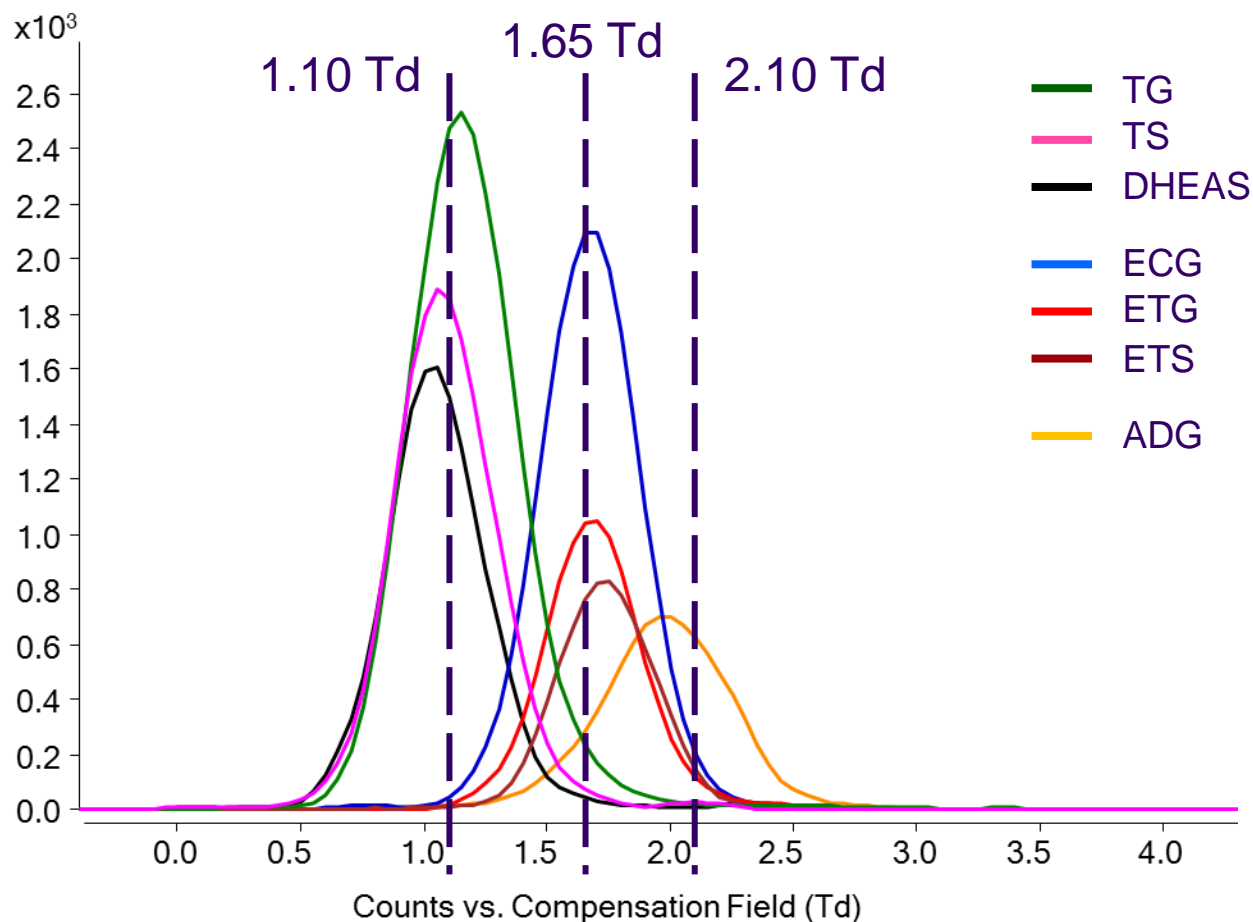


Testosterone glucuronide  
Epitestosterone glucuronide



Testosterone sulfate  
Epitestosterone sulfate

# FAIMS-MS of steroid metabolites



Dispersion Field 280 Td

FAIMS-MS in ACN:H<sub>2</sub>O (LC mobile phase)

# Separation of steroids using FAIMS-MS

| FAIMS+<br><i>m/z</i> | ETG | TG | ECG | ADG | DHEAS | TS | ETS |
|----------------------|-----|----|-----|-----|-------|----|-----|
| ETG                  |     |    |     |     |       |    |     |
| TG                   |     |    |     |     |       |    |     |
| ECG                  |     |    |     |     |       |    |     |
| ADG                  |     |    |     |     |       |    |     |
| DHEAS                |     |    |     |     |       |    |     |
| TS                   |     |    |     |     |       |    |     |
| ETS                  |     |    |     |     |       |    |     |



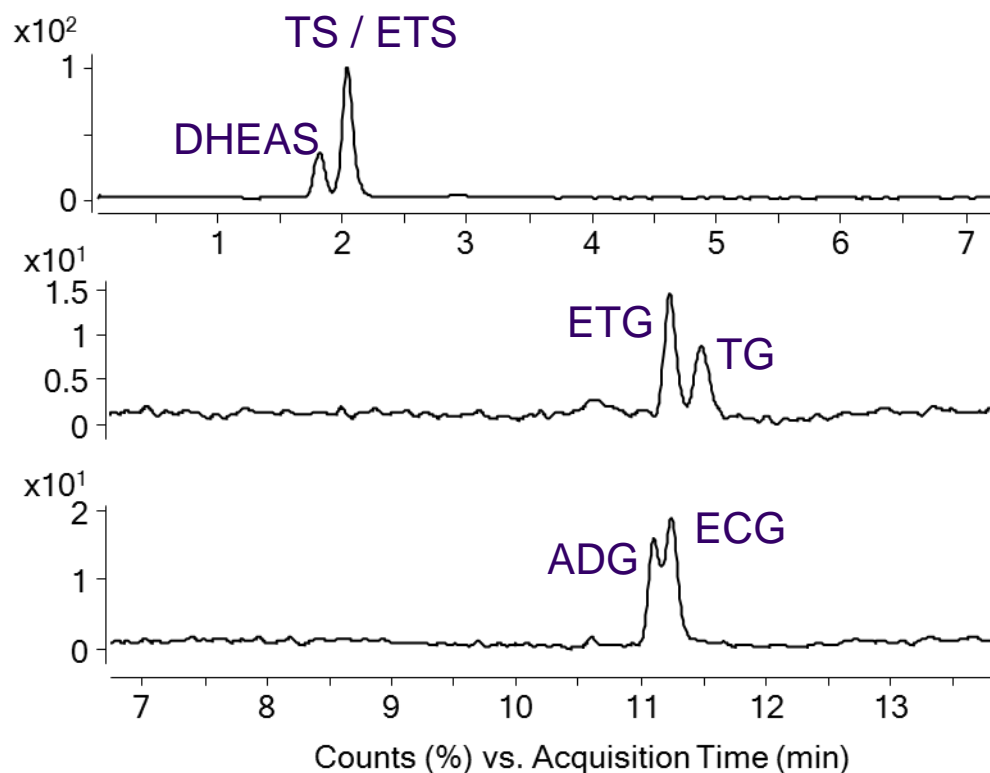
# HILIC of steroid metabolites

- DHEAS is not separated from TS by FAIMS or MS, so must be separated chromatographically using LC
- LC method should be a quick and reliable method for the determination of all of the compounds of interest in a urine matrix
- Hydrophilic interaction liquid chromatography (HILIC) was chosen as separation between DHEAS / TS could be achieved on a fast timescale



# LC-MS of steroids

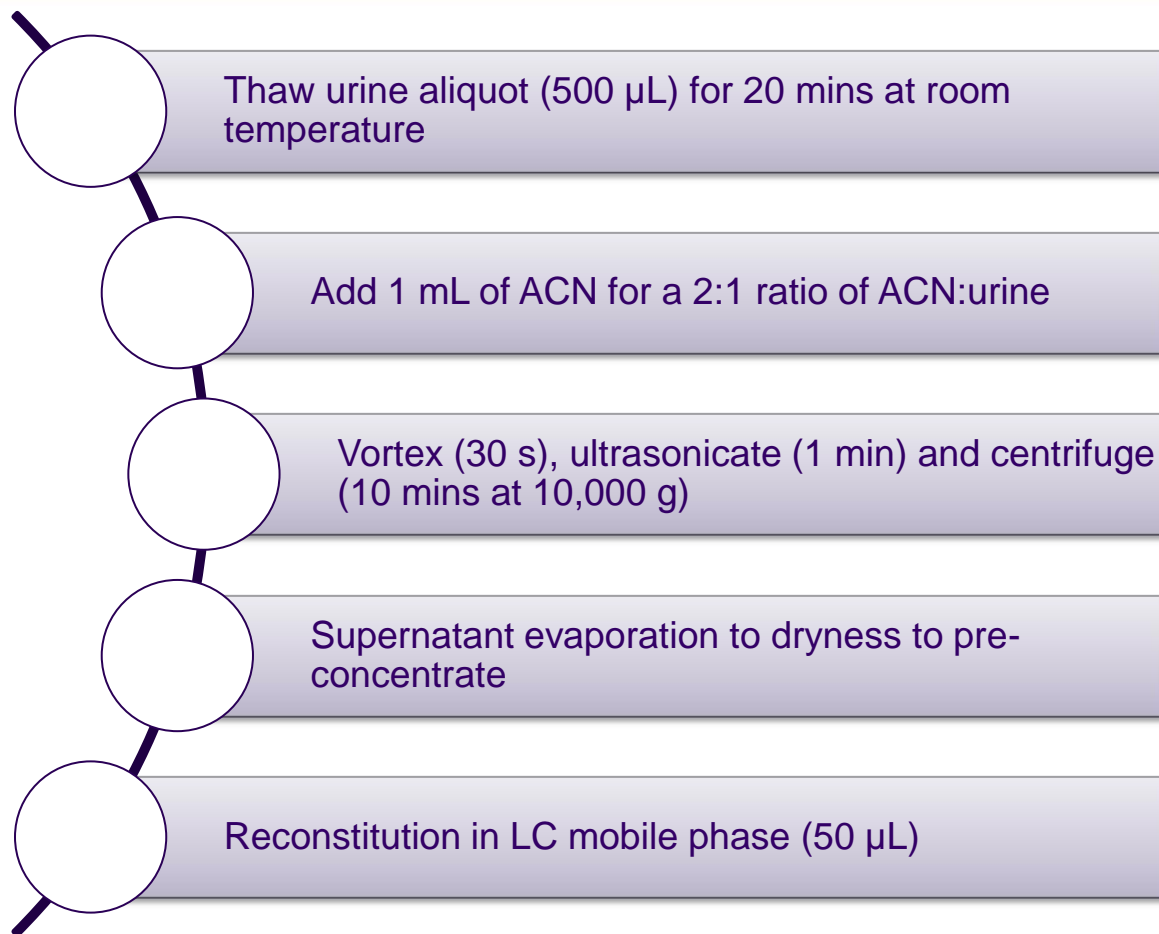
- Selected ion responses for  $m/z$ :
  - 413.14
  - 509.21
  - 511.23
- No chromatographic separation between TS / ETS
- ETG / TG are not baseline resolved
- Only partial separation between ADG / ECG



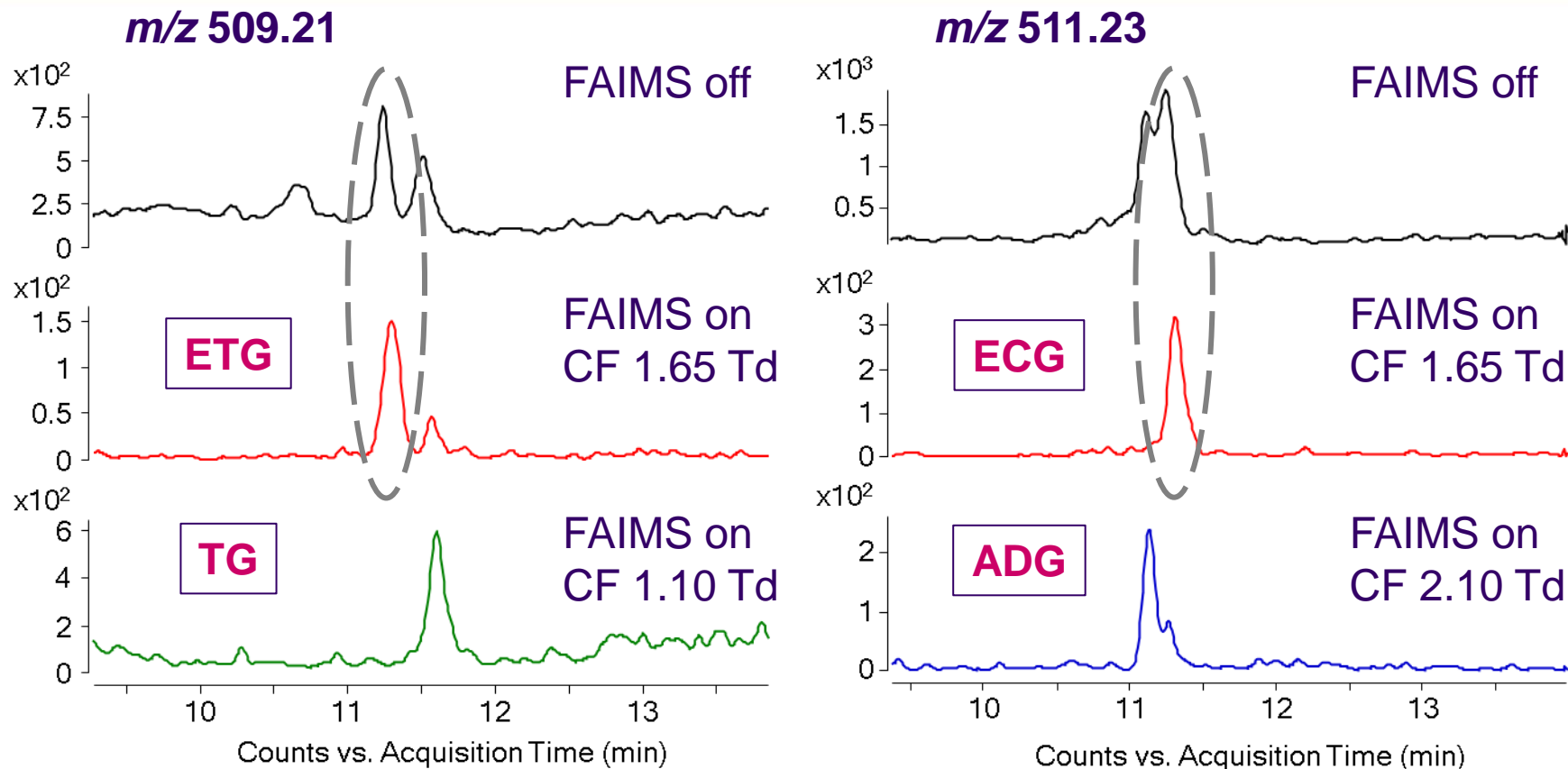
# Separation of steroids using LC-MS

| RT+ <i>m/z</i> | ETG | TG  | ECG   | ADG   | DHEAS | TS    | ETS   |
|----------------|-----|-----|-------|-------|-------|-------|-------|
| ETG            |     | Red | Green | Green | Green | Green | Green |
| TG             |     |     | Green | Green | Green | Green | Green |
| ECG            |     |     |       | Red   | Green | Green | Green |
| ADG            |     |     |       |       | Green | Green | Green |
| DHEAS          |     |     |       |       |       | Green | Green |
| TS             |     |     |       |       |       |       | Red   |
| ETS            |     |     |       |       |       |       |       |

# Urine sample preparation



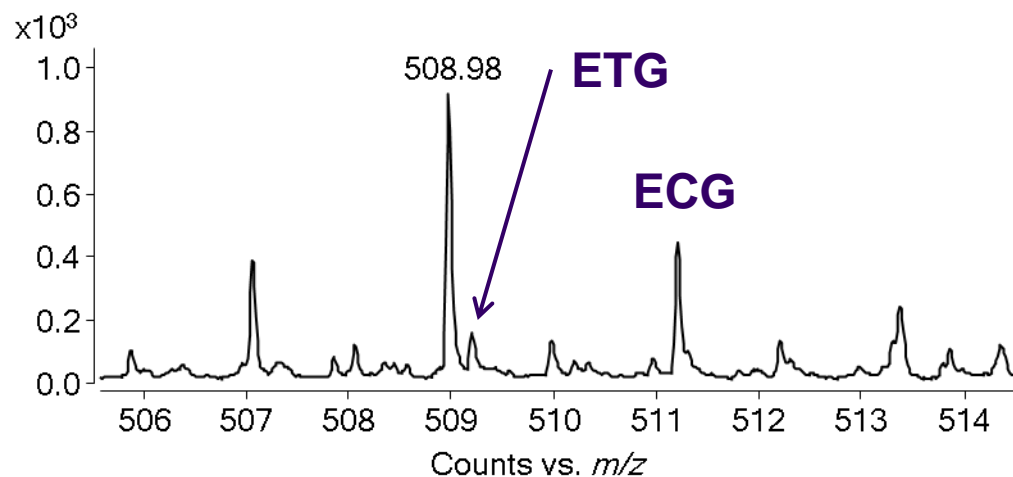
# LC-FAIMS-MS of steroids in urine



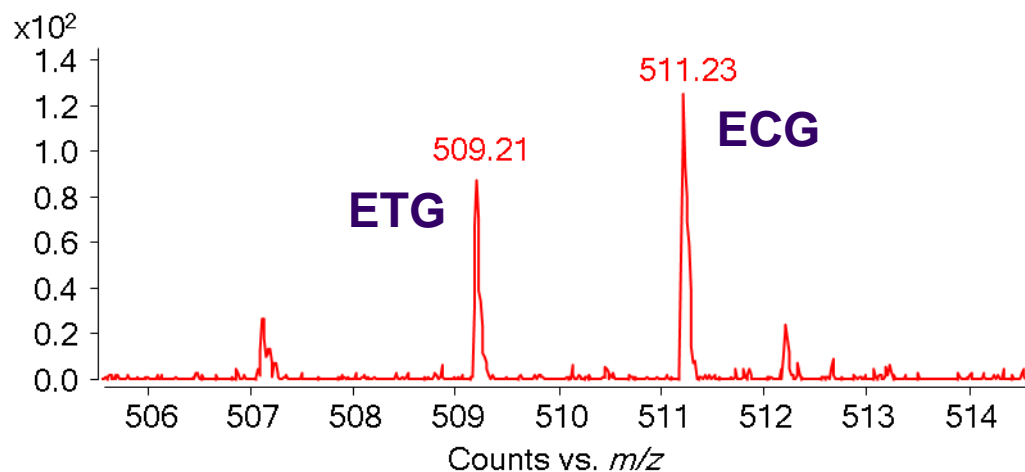
FAIMS on – Dispersion Field 280 Td

# LC-FAIMS-MS of steroids in urine

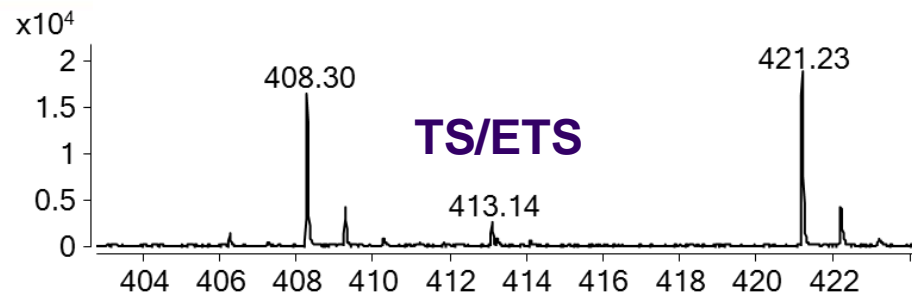
Mass spectra at RT 11.3 mins



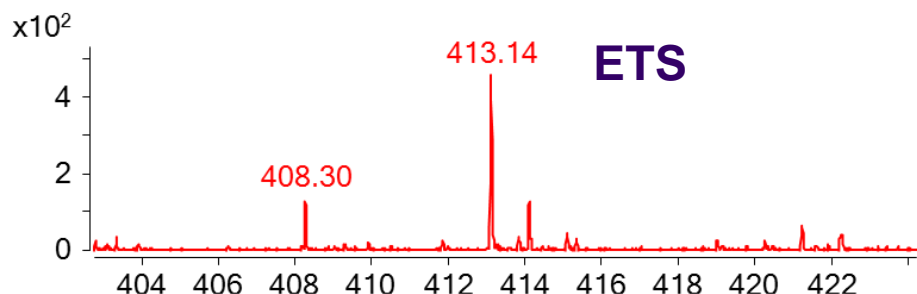
Dispersion Field 280 Td



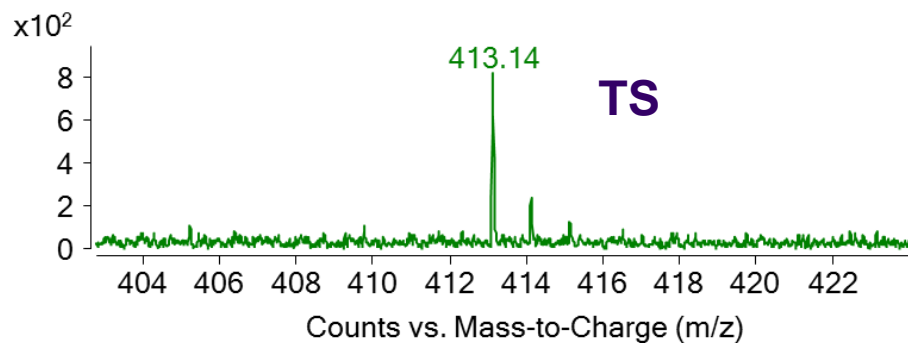
# LC-FAIMS-MS of steroids in urine



FAIMS off



FAIMS on  
CF 1.65 Td



FAIMS on  
CF 1.1 Td

Mass  
spectra at  
RT 2.2 mins

Dispersion Field  
280 Td

# Separation of steroids using LC-FAIMS-MS

| RT+ FAIMS<br>+ m/z | ETG | TG | ECG | ADG | DHEAS | TS | ETS |
|--------------------|-----|----|-----|-----|-------|----|-----|
| ETG                |     |    |     |     |       |    |     |
| TG                 |     |    |     |     |       |    |     |
| ECG                |     |    |     |     |       |    |     |
| ADG                |     |    |     |     |       |    |     |
| DHEAS              |     |    |     |     |       |    |     |
| TS                 |     |    |     |     |       |    |     |
| ETS                |     |    |     |     |       |    |     |



# Conclusion

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- FAIMS-MS separation of steroid sulfates and glucuronides in positive ionisation mode
- Improved FAIMS separation of steroids using sodiated adducts
- Potential to improve quantitative determination of steroids in biological matrices using LC-FAIMS-MS

# Further work

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- Further optimisation of method parameters
- Develop and optimise urine sample preparation with a pre-concentration step for maximum sensitivity
- Validation of the LC-FAIMS-MS method for urine analysis

# Acknowledgements

- **Loughborough University:**

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- Lauren Brown
- Robert Smith

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- David Cowan
- Andrew Kicman
- Alan Brailsford

- **Staff and researchers at the Centre for Analytical Science**

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[uk.linkedin.com/pub/kayleigh-arthur/8a/234/51](https://uk.linkedin.com/pub/kayleigh-arthur/8a/234/51)

